A 68 year-old male presented with an incidentally discovered gastric submucosal mass measuring 3.1 cm. A fine needle aspiration biopsy was performed and a diagnosis of "neoplasm consistent with a well-differentiated neuroendocrine tumor" was made. The resection specimen is demonstrated below.



Figure 1. Stomach, gross specimen: left-serosal surface; right-mucosal surface. On cut section the tumor was tan, lobulated, and well circumscribed (images not available).



Figure 2. Gastric neoplasm, H&E (1x magnification).



Figure 3. Gastric neoplasm, H&E (10x magnification).



Figure 4. Gastric neoplasm, H&E (20x magnification).



Figure 5. Left: Synaptophysin (1x magnification); Right: Chromogranin (1x magnification)



Figure 6. Left: Pan-cytokeratin (1x magnification); Right: CD34 (1x magnification)



Figure 7. Left: CD117 (1x magnification); Right: DOG1 (20x magnification)



Figure 9. Smooth Muscle Actin (1x magnification)

Other markers:

Vimentin – Positive Calponin – Positive Caldesmon – Patchy positive AE1/3 – Negative HMB45 – Negative Calretinin – Negative Inhibin – Negative Desmin – Negative S100 – Negative

Question: Based on the gross features, morphology, and immunophenotype, what is the most appropriate diagnosis?

- A) Well-differentiated neuroendocrine tumor
- B) Epithelioid gastrointestinal stromal tumor
- C) Glomus tumor
- D) Paraganglioma

Answer: C)

Discussion:

C) Glomus tumors (GTs) arise from specialized vascular smooth muscle cells (pericytes) and classically present as painful lesions on the subungual finger¹⁻². Glomus tumors of the stomach are uncommon lesions, which have been described in numerous case reports and series³⁻⁵. These tumors can clinically and morphologically mimic more common gastric neoplasms, and therefore knowledge of this entity, along with a high index of suspicion, are necessary for correct diagnosis³.

Clinically, patients with gastric GTs frequently present with epigastric pain, ulceration, or upper gastrointestinal bleeding, which can be severe³. Incidental presentations have also been reported³. A wide age range can be affected (median age 54 years), and a slight female predominance has been shown³. A wide variation in tumor size has been described, with a median size of 2.7 centimeters³. Larger gastric GTs may have an increased risk of metastasizing³.

Morphologically, GTs typically demonstrate well-circumscribed and sheet-like growth with prominent vascular spaces^{1,3}. The cells are usually monotonous and rounded with round-oval nuclei and eosinophilic to clear cytoplasm^{1,3}. Cell membranes are typically well-defined and the nuclei are centrally placed with delicate chromatin³. Correct diagnosis generally requires morphologic assessment along with a panel of immunohistochemical markers.

Glomus tumors of the gastrointestinal tract are usually positive for smooth muscle actin (SMA), calponin, and vimentin³. Peri-cellular net-like positivity is seen with Collagen type IV and Laminin³. One series found synaptophysin and CD34 positive in 18% and 20% of gastrointestinal glomus tumors, respectively³. Caldesmon is also positive in more than half of cases³. Cytokeratins, CD117 (c-kit), S100, desmin, chromogranin, and CD45 are consistently negative in these lesions³. Reports of DOG1 positivity in GTs (not restricted to the GI tract) have varied, with one series demonstrating 0/14 positive cases and another demonstrating 6/7 positive cases⁶⁻⁷. In the study by *Wong et al.* evaluating DOG1 positivity in numerous tumor types, glomus tumors were the only neoplasms to stain CD117 negative and DOG1 positive⁷.

Gastric GTs almost always follow a benign clinical course, however notable exceptions do exist in the literature. *Song et al.* reported the case of a 65 year-old woman presenting with a malignant gastric GT and multi-organ metastases⁸. In a case series of 32 patients, *Miettenen et al.* identified one patient with a gastric GT metastatic to the liver, who eventually died of their disease³. In another series by *Folpe et al*, the only case arising in the stomach had metastasized to the liver⁹.

In the study by *Folpe et al.*, they looked at 52 atypical and malignant GTs (from all body sites), and classified them as 1) malignant GT, 2) symplastic GT, 3) GTof uncertain malignant potential, and 4) glomangiomatosis⁸. This classification was based on features that included: size, site (superficial vs. deep), nuclear grade, presence of atypical mitotic figures, and mitotic rate⁸. Metastases were observed in 38% of tumors classified as malignant, and were not observed in the other three categories⁸. Of note, this classification scheme may not accurately predict the behavior of gastric GTs, which generally behave less aggressively than other deep-seated lesions, and may require a site-specific grading scheme³⁻⁴. Interestingly, BRAF V600E mutations have recently been identified in a subset of atypical/malignant GTs¹⁰.

A) One of the primary considerations in this case was a well-differentiated neuroendocrine tumor (NET). Morphologically, well-differentiated NETs have monotonous round-polygonal cells with round nuclei containing coarse 'salt and pepper' chromatin^{1, 7}. They are more commonly centered in the mucosa or submucosa, and architectural patterns can include nested, trabecular, and acinar^{1, 7}. By immunohistochemistry, well-differentiated NETs are typically positive for cytokeratins, CD56, chromogranin, and synaptophysin^{1, 7}. Lesions that are cytokeratin negative but synaptophysin and/or

chromogranin positive should prompt consideration for alternate diagnoses including GT or paraganglioma.

B) Another strongly considered differential in this case was an epithelioid gastrointestinal stromal tumor (GIST). Morphologically, epithelioid GISTs can also have clear cytoplasm and stromal hyalinization, similar to a GT³. However, they typically have more polygonal or oval cells, and do not usually have such prominent dilated veins or capillaries³. Among all GISTs (including spindled and epithelioid), roughly 60-70% will be positive for CD34, 95% will be DOG1 positive, and 95% will be CD117 positive. Both DOG1 and CD117 have similar sensitivities for diagnosing GISTs, and only approximately 1% of cases will be negative for both¹. While multiple different tumor types can express either CD117 or DOG1, only GISTs will show strong expression of both¹. However, it is important to remember that epithelioid GISTs can have markedly decreased or absent DOG1 and/or CD117 expression⁷. The majority of GISTs will have either KIT or PDGFRA mutations, which can be tested for in diagnostically challenging cases⁷.

d) Paragangliomas of the stomach are exceedingly rare tumors, though cases have been reported¹¹. Morphologically, extra-adrenal paragangliomas often show anastomosing cell cord or trabecular arrangements¹. Other variants include organoid (Zellballen) and solid/diffuse patterns¹. Cytoplasm is relatively abundant and finely granular, while nuclear pleomorphism is common¹. By immunohistochemistry, paragangliomas are usually positive for neuroendocrine markers, including synaptophysin and chromogranin, and negative for cytokeratins (though not always)^{1, 3}. S100 positive sustentacular cells are another helpful clue to making the diagnosis^{1, 3}.

Key Points:

- Glomus tumors of the stomach are uncommon lesions that can mimic more common tumors, especially on small biopsy specimens. A high index of suspicion is required.
- These tumors can be positive for synaptophysin but should always be negative for cytokeratins; a feature which can help to differentiate them from well-differentiated neuroendocrine tumors.
- Glomus tumors can also be positive for DOG1 and CD34, potentially mimicking an epithelioid gastrointestinal stromal tumor (GISTs). However, CD117 should be negative. PDGFRA or KIT mutation testing can be performed in difficult cases.
- Most gastric glomus tumors follow a benign clinical course, however notable exceptions do exist in the literature. Features that include increased size and mitotic rate may predict aggressive behavior.

References:

1. Mills, S. E., *et al.* (2015). Sternberg's diagnostic surgical pathology: Sixth edition. Wolters Kluwer Health Adis (ESP).

2. Mosquera, J-M., *et al.* (2013) Novel MIR143-NOTCH fusions in benign and malignant glomus tumors. *Genes, Chromosomes & Cancer*, 52, 1075-1087.

3. Miettinen, M., *et al.* (2002). Gastrointestinal glomus tumors: a clinicopathologic, immunohistochemical, and molecular genetic study of 32 cases. *American Journal of Surgical Pathology*, 26(3), 301–311.

4. Kang, G., *et al.* (2012). Glomus tumor of the stomach: A clinicopathologic analysis of 10 cases and review of the literature. *Gut and Liver*, *6*(1), 52–57.

5. Bennett, S., *et al.* (2015). A case series of two glomus tumors of the gastrointestinal tract. *Journal of Surgical Case Reports*, (1), 1-3.

6. Miettinen, M., *et al.* (2009). DOG1 Antibody in the Differential Diagnosis of Gastrointestinal Stromal Tumors: A Study of 1840 Cases. *American Journal of Surgical Pathology*, 33(9), 1401–1408.

7. Wong, N. A. C. S., *et al.* (2010). Specificity of DOG1 (K9 clone) and protein kinase C theta (clone 27) as immunohistochemical markers of gastrointestinal stromal tumour. *Histopathology*, *57*(2), 250–258.

8. Song, S. E., *et al.* (2010). Malignant glomus tumor of the stomach with multiorgan metastases: Report of a case. *Surgery Today*, *40*(7), 662–667.

9. Folpe, A. L., *et al.* (2001). Atypical and malignant glomus tumors: analysis of 52 cases, with a proposal for the reclassification of glomus tumors. *American Journal of Surgical Pathology*, 25(1), 1–12.

10. Karamzadeh, D. N., *et al.* (2017). BRAF V600E Mutations Occur in a Subset of Glomus Tumors, and are Associated With Malignant Histologic Characteristics. *American Journal of Surgical Pathology*, *41*(11), 1532-1541.

11. Bura, R., *et al.* (2014). Gastric paraganglioma: a case report and a review of the literature. Annali Italiani Di Chirurgia, 85(ePub), 84–89.

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